Action: respond to our copy-editing questions

Select each question and describe any changes we should make on the proof. Changes against journal style will not be made and proofs will not be sent back for further editing.

- AQ1. Please check all author names and affiliations. Please check that author surnames have been identified by a pink background. This is to ensure that forenames and surnames have been correctly tagged for online indexing.
- AQ2. If your manuscript has figures or text from other sources, please ensure you have permission from the copyright holder. For any questions about permissions contact <u>inls.author.support@oup.com</u>.
- AQ3. Please check that funding is recorded in a separate funding section if applicable. Use the full official names of any funding bodies, and include any grant numbers.
- AQ4. You may need to include a "conflict of interest" section. This would cover any situations that might raise any questions of bias in your work and in your article's conclusions, implications, or opinions. Please see <u>here</u>.
- AQ5. Please provide the department/unit (if any) for affiliations 3–5.
- AQ6. Kindly check and confirm heading levels.
- AQ7. Please review the typeset table carefully against copies of the originals to verify accuracy of editing and typesetting.
- AQ8. Figures have been placed as close as possible to their first citation. Please check that they have no missing sections and that the correct figure legend is present.
- AQ9. The following references are not included in the reference list: "Oksanen et al. 2013; McKechnie and Wolf 2010; Orr et al. 2014". Please include the citations in the list with full publication details.
- AQ10. The following references are not cited in the main text: Howlett 1996, Mahoney 2020, Strong 1990, Yamaura 2019. Please cite them in the text or delete from the list.
- AQ11. DOI number given for the reference "Mahoney et al. 2022" seems not correct. Please check and amend as necessary.

These proofs are for checking purposes only. They are not in final publication format. Please do not distribute them in print or online. Do not publish this article, or any excerpts from it, anywhere else until the final version has been published with OUP. For further information, see https://academic.oup.com/journals/pages/authors

Figure resolution may be reduced in PDF proofs and in the online PDF, to manage the size of the file. Full-resolution figures will be used for print publication.

Action: check your manuscript information

Please check that the information in the table is correct. We use this information in the online version of your article and for sharing with third party indexing sites, where applicable.

Full affiliations Each unique affiliation should be listed separately; affili- ations must contain only the applicable department, insti- tution, city, territory, and country	 Department of Biological Sciences, Northern Arizona University, Flagstaff, Arizona, USA Department of Biological Sciences, Thompson Rivers University, Kamloops, British Columbia, Canada Colorado Plateau Research Station, Northern Arizona University, Flagstaff, Arizona, USA Marine Science Institute, University of California, Santa Barbara, Santa Barbara, California, USA Bloom Research Inc., Santa Ana, California, USA
Group Contributors The name of the group and individuals in this group should be given, if applicable (e.g. The BFG Working Group: Simon Mason, Jane Bloggs)	NA
Supplementary data files cited	
Funder Name(s) Please give the full name of the main funding body/agency. This should be the full name of the funding body without abbreviation or translation, if unsure, see https://search. crossref.org/funding	the Landscape Conservation Cooperative (LCC) Clark County Multiple Species Habitat Conservation Plan (MHSCP) the United States Department of Interior (USDI)

How to add your responses

These instructions show you how to add your responses to your proof using Adobe Acrobat Professional version 7 onwards, or Adobe Reader DC. To check what version you are using, go to 'Help', then 'About'. The latest version of Adobe Reader is available for free from https://get.adobe.com/uk/reader/.

Displaying the toolbars

Adobe Reader DC

In Adobe Reader DC, the Comment toolbar can be found by clicking 'Comment' in the menu on the top-right-hand side of the page (shown below).



The toolbar shown below will then display along the right-hand-side of the page.

						×
	Тоо	ls	Sig	jn	Con	nment
	▼ Ann	otat	tions			
	ج (J	Т	G	4	-
	T _e	F	Ŧ	<u>T</u>	Ъ	T☆
	▼ Dra	wing	g Mar	kups		
	Ţ	Þ	_	⇒	\bigcirc	
	\bigcirc	\bigcirc	Ø	Ø	0	
	▼ Con	ıme	nts Li	st (0)		
	🔍 Fin	d			Å-	≥ - 8=-
	This	docu	iment	has n	o com	ments.
1						

Acrobat Professional 7, 8 and 9

In Adobe Professional, the Comment toolbar can be found by clicking 'Comment(s)' in the top toolbar, and then clicking 'Show Comment & Markup Toolbar' (shown below).

🔗 Comment 🔹	
😑 Add Sticky <u>N</u> ote	Ctrl+6
Show Comment & Markup Toolbar	N
🖇 Show Comments <u>L</u> ist	13
🔁 Attach for Email Review	
and for Shared Review	
🚰 Trac <u>k</u> Reviews	

The toolbar shown below will then be displayed along the top of the page.



Using text edits and comments in Acrobat

This is the easiest method to both make changes, and for your changes to be transferred and checked.

- 1. Click 'Text Edits'
- 2. Select the text to be annotated or place your cursor at the insertion point and start typing.
- 3. Click the 'Text Edits' drop down arrow and select the required action.
- 4. You can also right click on selected text for a range of commenting options, or to add sticky notes.



Using commenting tools in Adobe Reader

All commenting tools are displayed in the toolbar. You cannot use text edits, however you can still use highlighter, sticky notes, and a variety of insert/replace text options.



Pop-up notes

In both Reader and Acrobat, when you insert or edit text, a pop-up box will appear.

Saving comments

In order to save your comments and notes, you need to save the file ('File', 'Save') before closing the document.

NB: Do not make any edits directly into the text, use commenting tools only

	Applications	AmericanOrnithology.org	
		Volume XX, 2022, pp. 1–16 https://doi.org/10.1093/ornithapp/duac012	1.55
	RESEARCH ARTICLE		
1.5	Tamarisk biocontrol alters bird community co cottonwood and willow vegetation	mposition in the absence of	AQ1-AQ4 1.60
	Sean M. Mahoney, ^{1,2,*,®} Matthew J. Johnson, ³ Jennifer A. Holmes, ³	[*] Tom L. <mark>Dudley</mark> , ⁴ Michael <mark>Kuehn</mark> , ^{4,5} and	
1.10	 ¹ Department of Biological Sciences, Northern Arizona University, Flagstaff, Arizon ² Department of Biological Sciences, Thompson Rivers University, Kamloops, Briti ³ Colorado Plateau Research Station, Northern Arizona University, Flagstaff, Arizon ⁴ Marine Science Institute, University of California, Santa Barbara, Santa Barbara, Santa Barbara, Santa Ana, California, USA *Corresponding author: sm2275@nau.edu 	na, USA sh Columbia, Canada na, USA California, USA	1.65 _{AQ5}
1.15	Submission Date: November 18, 2021; Editorial Acceptance Date: March 1, 2022;	Published: Month 00, 2022	4 = 0
	Submission Date: November 17, 2021; Editorial Acceptance Date: March 11, 2022; Publisher	d March 12, 2022	1.70
1.20	ABSTRACT Invasive plants threaten biodiversity worldwide, but control of non-nat plex ways. Non-native tamarisk (<i>Tamarix</i> spp.) is widespread in western U. (<i>Diorhabda</i> spp.), a tamarisk-specific herbivore, were subsequently introc control is defoliation and branch dieback, with repeated defoliation killir	ive species may affect native species in com- S. riparian environments, and tamarisk beetles duced as biocontrol. The primary effect of bio- ng the plant. We investigated the initial stages	1.75
1.25	of site recovery after biocontrol and how tamarisk decline affected bird compared avian community diversity and composition, arthropod abund the Virgin River in Nevada and Arizona, USA characterized as either mix tamarisk-dominated where the majority of tamarisk died from biocontro after biocontrol to counts at the same locations before biocontrol. Prior but one site grouped together using unbiased clustering algorithms. For	s, their arthropod prey, and microclimate. We ance, humidity, and temperature at sites along ed native vegetation, tamarisk-dominated, or ol. We compared avian communities sampled to biocontrol, community compositions of all ollowing biocontrol, tamarisk-dominated sites	1.80
1.30	grouped separately, and mixed sites grouped with the pre-biocontrol cli communities showed 7 common species declined by ≥30% in dead tamar and 3 at tamarisk-dominated sites. Individual census points in dead tamar than the same points censused before biocontrol, unless native vegetation the cause of dominant species abundance changes. Tamarisk-dominated tive vegetation and supported fewer non-tamarisk-obligate arthropods, of tions were driven by changes in microclimate and prey abundance. How lo	uster. Comparison of pre- and post-biocontrol isk sites, while one species did so at mixed sites arisk had significantly lower Simpson diversity on was present, suggesting tamarisk death was sites were hotter and drier than sites with na- consistent with the hypothesis that bird reduc- ong these effects last will depend upon the rate	1.85
1.35	of native vegetation recovery after biocontrol, therefore we recommend a vegetative recovery and considering the need and feasibility of active re regeneration. <i>Keywords:</i> biological control, bird communities, defoliation, riparian, ta	monitoring sites to determine the trajectory of estoration in those sites with slow or no native amarisk, tamarisk beetles, Tamarix	1.90
1.40	LAY SUMMARY		1.95
1 45	 Invasive species disturb ecosystems and threaten biodiversity. Invasive species disturb ecosystems and threaten biodiversity. Invasive species control, can cause additional disturbances, so quantifying how native species to inform best management practices We quantified southwestern bird communities in sites that varied in the 	species management, such as biological becies respond to invasive control is important e amount of the non-native plant tamarisk	1.75
1.10	 (Tamarix spp.), before and after biological control efforts Following biocontrol, we found significant differences in community co species declined by ≥30% Bird declines were ameliorated in the presence of native vegetation, control of the presence of the pr	mposition and diversity, and several bird nsistent with the hypothesis that tamarisk	1.100
1.50	 biocontrol decreases prey availability and alters microclimate We recommend land managers monitor areas dominated by tamarisk a 	fter biocontrol, and if re-establishment of	

We recommend land managers monitor areas dominated by tamarisk after biocontrol, and if re-establishment of
native vegetation is slow or lacking, consider the feasibility of active restoration

1.105

1.106

1.54

Ornithological

2.10

El control biológico del tamarisco altera la composición de la comunidad de aves en ausencia de vegetación de álamos y sauces	2.55
RESUMEN	
Las plantas invasoras amenazan la biodiversidad en todo el mundo, pero el control de las especies no nativas puede afectar a las especies nativas de formas complejas. El tamarisco (<i>Tamarix</i> spp.), una especie no nativa, está muy extendido en los ambientes ribereños del oeste de los EEUU, y el escarabajo del tamarisco (<i>Diorhabda</i> spp.), un herbívoro específico del tamarisco, fue introducido posteriormente como control biológico. El efecto principal del control biológico es la defoliación y el secado de las ramas, haciendo que la defoliación repetida mate la planta. Investigamos las etapas	2.60
iniciales de la recuperación del sitio después del control biológico y cómo la disminución del tamarisco afectó a las aves, sus presas artrópodas y el microclima. Comparamos la diversidad y composición de la comunidad de aves, la abundancia de artrópodos, la humedad y la temperatura en sitios a lo largo del Río Virgin en Nevada y Arizona, EEUU, caracterizados como vegetación nativa mixta, dominada por tamariscos, o dominada por tamariscos donde la mayoría de los tamariscos murió a causa del control biológico. Comparamos las comunidades de aves muestreadas después del control biológico con conteos realizados en los mismos sitios antes del control biológico. Antes del control biológico,	2.65

las composiciones de las comunidades de todos los sitios menos uno se agruparon juntas utilizando algoritmos de agrupamiento no sesgados. Después del control biológico, los sitios dominados por tamariscos se agruparon por 2.15 separado y los sitios mixtos se agruparon con el grupo previo al control biológico. La comparación de las comunidades 2.70 antes y después del control biológico mostró que siete especies comunes disminuyeron en ≥30% en los sitios de tamariscos muertos, mientras que una especie lo hizo en sitios mixtos y tres en sitios dominados por tamariscos. Los puntos de censo individuales en tamariscos muertos tuvieron una diversidad de Simpson significativamente menor que los mismos puntos censados antes del control biológico, a menos que hubiera vegetación nativa presente, lo que 2.20 sugiere que la muerte de los tamariscos fue la causa de los cambios en la abundancia de las especies dominantes. Los sitios dominados por tamariscos fueron más cálidos y secos que los sitios con vegetación nativa y albergaron menos 2.75artrópodos no obligados de los tamariscos, lo que concuerda con la hipótesis de que la reducción de las aves estuvo impulsada por cambios en el microclima y en la abundancia de presas. La duración de estos efectos dependerá de la tasa de recuperación de la vegetación nativa después del control biológico; por lo tanto, recomendamos monitorear los sitios para determinar la trayectoria de la recuperación vegetativa y considerar la necesidad y viabilidad de la restauración 2.25activa en aquellos sitios con regeneración nativa lenta o nula. 2.80

Palabras clave: comunidades de aves, control biológico, defoliación, escarabajo del tamarisco, ribereño, tamarisco, Tamarix

2.30 INTRODUCTION

Non-native plant introductions may alter the structure, composition, and function of habitat, potentially changing the relative value of the habitat for the native faunal community (Vilà et al. 2011). Globally, non-native spe-2.35 cies are becoming increasingly widespread in introduced ranges, and may interact with native faunal communities in a variety of ways (reviewed in Rodriguez 2006, Hobbs et al. 2009, 2018), so management of non-native introductions has become an important goal in conservation 2.40biology. Classical biological control (hereafter, biocontrol) is a common cost-effective approach to manage naturalized non-native species. Introductions of biocontrol may also interact with native species in complex ways (Pearson and Callaway 2003); therefore, quantifying the responses 2.45of native species to non-native biocontrol is important to understand implications for the broader ecological community and to inform best management practices.

In the western United States, non-native tamarisk (*Tamarix* spp., also known as saltcedar), is now the third most dominant woody riparian plant (Friedman et al. 2005). To manage the continued spread of tamarisk, a biological control agent, the tamarisk beetle (*Diorhabda* spp.), an herbivorous tamarisk-obligate insect from southern Europe and North Africa that consumes tamarisk foliage, was introduced into the United States in 2001 (Bean et al. 2.85 2013). Since the release, tamarisk beetles have defoliated thousands of hectares of tamarisk in the western U.S. (Sogge et al. 2008, Dudley and Bean 2012, Nagler et al. 2012). The tamarisk beetle was expected to colonize new areas slowly due to physiological diapause limitations (DeLoach et al. 2000, Lewis et al. 2003, USDA 2005), but in roughly ten generations tamarisk beetles adapted and established populations across a broader geographic area than anticipated (Bean et al. 2013).

Tamarisk spread and subsequent biocontrol is a con-2.95 servation concern for native bird communities because although only 1% of the western USA landscape is riparian habitat, these areas support disproportionately more breeding birds than do surrounding upland habitats (Johnson et al. 1977, Stevens et al. 1977, Skagen et al. 1998, 2.100 Cartron et al. 1999). The effect of tamarisk invasion on na-AQ6 tive riparian birds is complex depending on geographic region, climate, and stand composition, and it is confounded by variation in climate and stand composition. A variety of birds will use and breed in tamarisk (Hunter et al. 1988, 2.105 Sogge et al. 2008, 2013), but several studies have documented lower avian abundance and richness in tamarisk relative to native Fremont cottonwood (Populus fremontii) 2.106

3.10

3.15

3.54

S. M. Mahoney et al.

and willow (*Salix* spp.) stands, and in areas where high summer temperatures may preclude nesting in tamarisk because it lacks the multi-layered canopy cover of native stands (Anderson and Ohmart 1977, Ellis 1995, Brand et al. 2008). Others have argued that total vegetation volume and structural diversity drives richness and diversity independent of the identity of the dominant tree (Fleishman et al. 2003, Cable et al. 2015, Raynor et al. 2017). A confounding factor in studies evaluating the role of tamarisk as habitat for native birds may be variation across study sites in the abundance of tamarisk relative to native vegetation, as use by native birds may depend upon the presence and relative abundance of native trees within a stand (Holmes et al. 2005, Sogge et al. 2008, van Riper et al. 2008; and reviewed in Shafroth et al. 2005).

Much less is known about avian responses to biocontrol of tamarisk. Biocontrol could affect birds positively in the short term by increasing prey abundances for bird species that include tamarisk beetles and their larvae in the diet, and in the long term if biocontrol is successful in allowing 3.20 native riparian vegetation to replace tamarisk (Paxton et al. 2011). The potential for tamarisk beetles to serve as prey for birds is supported by studies from tamarisk-dominated riparian areas of northern Nevada that documented higher indices of bird use and greater avian diversity in tamarisk 3.25 stands in which tamarisk beetles and their larvae were present compared to stands lacking beetles (Longland and Dudley 2008, Bean et al. 2013). In contrast, fecal diet analyses of native insectivorous birds in southern Nevada (Virgin River) and Colorado (Dolores River) found tam-3.30 arisk beetles were not selected (Mahoney et al. 2017, van Riper et al. 2018). Alternatively, defoliation caused by the beetle could decrease abundance of other insect prey. For example, tamarisk leafhoppers (Opsius stactogalus), a tamarisk-obligate insect also found in diets of several na-3.35 tive birds (Yard et al. 2004, Wiesenborn and Heydon 2007, Durst et al. 2008, Mahoney et al. 2017, van Riper et al. 2018), declined significantly when tamarisk defoliation reached 60% (Eckberg and Rice 2016). Defoliation by the beetle could also reduce nesting cover while increasing 3.40 temperature and reducing humidity due to increased solar radiation. Only one study we are aware of reported avian productivity in sites defoliated by tamarisk beetles and it found decreased nest success by Southwestern Willow 3.45 Flycatchers (Empidonax trailii extimus) in defoliated habitats, with nest predation and failure to hatch as the major causes (McLeod 2018). Failure to hatch was rare at these sites in other years, and at other sites during the same year, but was consistent with eggs being addled due to higher 3.50 nest temperatures (McLeod 2018).

> In this study, we compared avian communities sampled in 2009–2010 before the effects of biocontrol to the same sites sampled in 2013–2014 after extensive areas at some sites were defoliated as a result of tamarisk beetle

herbivory. During both sampling periods, sites were 3.55 characterized as either mixed native vegetation (50-75% of canopy comprised of native vegetation), or tamariskdominated vegetation (70-90% canopy was tamarisk), allowing a comparison of sites differing in the amount of tamarisk, while in the later sample tamarisk-dominated 3.60 sites could be separated into those in which tamarisk were defoliated but not dead as a result of beetle herbivory compared to sites in which most tamarisk were dead or dying. We analyzed avian community responses to vegetation change at the landscape scale by comparing communities 3.65 across sites and at a smaller local scale more representative of individual territory size by comparing individual census points that differed in the presence or absence of a native plant component. At the larger spatial scale of sites, we predicted bird communities would be more diverse in 3.70 sites with a mix of native and tamarisk vegetation than in sites dominated by tamarisk prior to biocontrol. After biocontrol, we predicted avian community diversity would be lower in tamarisk-dominated sites that experienced dieback and death due to defoliation by the tamarisk beetle. At 3.75 smaller spatial scales, we predicted that diversity at census points with some native vegetation would be higher than at points that lacked a native component. Finally, although we lacked pre-biocontrol data, we predicted that after biocontrol, arthropod abundance would be lower, temperature 3.80 higher and humidity lower in tamarisk-dominated sites that experienced dieback and death due to loss of foliage.

METHODS

Study Area

Bird communities were assessed using point counts at 8 sites along the Virgin River in Arizona and Nevada, USA in 2009 and 2010 (prior to when beetles first arrived at the sites) and again in 2013 and 2014 after tamarisk beetles had been pre-3.90 sent for several years (Bateman et al. 2013) (Figure 1, Table 1). Sites varied in the proportion of tamarisk ("tamarisk" as used here refers to Tamarix ramosissima and related species and hybrids; Gaskin and Schaal 2002, Bateman et al. 2013) and 3.95 native vegetation (mainly Fremont cottonwood, Goodding's willow [Salix gooddingii], and coyote willow [Salix exigua]). In 2013, 2 observers independently estimated percentage of tamarisk and native riparian vegetation cover from a high point overlooking each site and the 2 estimates were averaged to obtain an index of vegetation type (Mahoney et al. 2017). 3.100 Sites with riparian vegetation cover comprising predominantly native cottonwood and willow vegetation (50-75% native) were categorized as "mixed native sites" (Beaver Dam, AZ [25% tamarisk cover], Mesquite, NV [40% tamarisk cover], Bunkerville, NV [50% tamarisk cover]); sites with 3.105 >75% tamarisk cover were categorized as "predominately defoliated tamarisk" sites ("tamarisk*" in figures and tables; Desert Springs, AZ [75% tamarisk cover], Big Bend, AZ [80% 3.106

4 Tamarisk biocontrol and native bird communities

S. M. Mahoney et al.

4.75

4.100

4.5

4.10

4.15

4.54



4.20 **FIGURE 1.** Invasive tamarisk and its introduced biocontrol are widespread throughout the Virgin River in Arizona and Nevada. Inset shows study area within the United States. Biocontrol at "Tamarisk*" sites defoliated but did not kill the plant, whereas biocontrol re-

tamarisk cover], Littlefield, AZ [80% tamarisk cover]); and sites with >75% tamarisk, but with the majority of the tam-4.25arisk at the site dead from repeated defoliation events, was categorized as "predominately dead tamarisk" sites ("tamarisk^" in figures and tables; Mormon Mesa 1, NV [90% dead tamarisk] and Mormon Mesa 2, NV [90% dead tamarisk]). During surveys in 2009 and 2010, no tamarisk experienced 4.30defoliation, but tamarisk at all sites in 2013 and 2014 during our surveys were being actively defoliated or had already been killed by beetles (Mosher and Bateman 2016). In our study we define "defoliation" as the response of tamarisk actively experiencing herbivory, including browning of foliage and die-4.35 back of some of the canopy (Hultine et al. 2015). We define "dead tamarisk" as those trees that had no foliage following biocontrol, with the caveat that some tamarisk may have resprouted in subsequent years but showed no evidence of doing so during the course of our study (Hultine et al. 2015). 4.40Our estimates of the status of tamarisk ("defoliated" or "dead") at our tamarisk-dominated sites were consistent with assessments by other researchers working independently at these same sites at the same time (Bateman et al. 2013, Hultine et al. 2015, Nagler et al. 2018). Native arrowweed (*Pluchea sericea*) was documented as increasing in abundance after tamarisk 4.45 biocontrol along the Virgin River (González et al. 2020), but because no point count locations (see below) were in areas dominated by arrowweed, we did not quantify its abundance.

4.50 Bird Species Diversity, Richness, and Community Composition

To assess the native bird community at each site, we conducted point-count surveys at 6 point locations within each site prior to the arrival of beetles in 2009 and 2010, and after beetle arrival in 2013 and 2014 following methods in Reynolds et al. (1980). Each point count location was separated by ~250 m. We visited each point 2 times during the 4.80breeding season (once in late May/June and once in July) each year. During each point count, we recorded all birds seen or heard during an 8-min period. We recorded the distance to the bird using laser range finders. To avoid over sampling, we only recorded individuals once during the 4.85 survey period. All surveys were conducted on clear days with low wind (≤ 3 on Beaufort scale or ≤ 20 km hr⁻¹) between sunrise and 1100 hours. Because all sites included some tamarisk and some native vegetation, and because tamarisk beetles were present at all sites, individual sam-4.90 pling points within any of the sites could potentially be located in areas of dead or defoliating tamarisk and/or have native vegetation present. Therefore, once per year in 2013–2014, the vegetation surrounding each point was categorized as 1 mix of green tamarisk and native vegeta-4.95 tion, 2 mix of defoliating tamarisk and some native vegetation, 3 mix of dead tamarisk and some native vegetation 4 green tamarisk, 5 defoliating tamarisk, 6 dead tamarisk, or 7 edge, in which any of the preceding categories was adjacent to open water, wetland, or upland.

Arthropod Surveys and Microclimate Estimates

During June and July 2013 and 2014, we repeatedly moni-
tored (n = 3-4 visits per point) the presence of tamarisk
beetles and other arthropods by sweeping vegetation
using canvas nets 5 times every 5 m along a 20-m tran-
sect (25 sweeps total per transect) at a subset of point
count locations, chosen arbitrarily (*sensu* Mahoney et al.
2017, Smith et al. 2017). Vegetation that was swept along4.1054.106

Ornithological Applications XX:1-16 © 2022 American Ornithological Society

Cop	vedited	bv:	OUP

5.10

5.15

5.20

5.25

5.30

5.35

5.40

5.45

5.50

5.54

S. M. Mahoney et al.

nunity composition c										
							Native	Tamarisk	Pre-biocontrol	Post-biocontrol
	State	Habitat	Area	ч	Latitude	Longitude	vegetation (%)	vegetation (%)	cluster	cluster
Dam (BD)	AZ	Mixed	25.4	9	36.89	-113.93	75	25	1	1
e (MQ1)	N	Mixed	29.5	9	36.79	-114.08	60	40	-	-
ille (MQ2)	N	Mixed	31.6	9	36.78	-114.13	50	50	1	1
prings (DS)	AZ	Tamarisk*	33.8	9	36.90	-113.89	25	75	2	2
d (BB)	AZ	Tamarisk*	47.8	9	36.84	-113.98	20	80	-	2
Id (LI)	AZ	Tamarisk*	44.2	9	36.83	-113.98	20	80	-	2
n Mesa 1 (MM1)	NV	Tamarisk∧	52.2	9	36.62	-114.32	10	90	-	2
(MM2) Mesa 2	N	Tamarisk∧	70.2	Ś	36.59	-114.33	10	00	-	2

transects included: Fremont cottonwood, coyote willow, 5.55 Goodding's willow, quailbush (*Atriplex lentiformis*), mesquite (*Prosopis* spp.), arrowweed, seepwillow (*Baccharis salicifolia*), cattail (*Typha* spp.), and tamarisk. We recorded the total numbers of 3 tamarisk-obligate insects (tamarisk leafhoppers, tamarisk weevils (*Coniatus splendidulus*), 5.60 and tamarisk beetles), as well as the number and type of any other arthropods (mainly spiders (Araneae), ladybird beetles (Coccinellidae) and ants (Formicidae)) collected in each sweep.

To assess how site temperature and relative humidity 5.65 were affected by tamarisk defoliation, in 2013 and 2014 we arbitrarily (*sensu* Smith et al. 2017) placed 2–4 microclimate dataloggers (HOBO model Pro v2) at a subset (n = 5) of native sites (Beaver Dam), mixed sites (Mesquite), defoliated tamarisk sites (Big Bend, Desert Springs), and dead tamarisk sites (Mormon Mesa 1). Microclimate dataloggers were placed 1.5-m high in an individual tamarisk, close to the trunk on 1 June and were retrieved on 31 July and recorded temperature and relative humidity every 30 min from 0600 to 1800 hours. We did not control for the cardinal direction of the logger on the trunk.

Data Analysis

Bird species diversity, richness, and community composition. We assessed species diversity by first combining 5.80 species detections across all 4 visits (1 June and 1 July visit in each of 2 years) at each sampling point in 2009-2010 and again in 2013-2014 to generate a list of all species detected at each point either pre- or post-biocontrol. With 6 sampling points at each site, this generated 18 points in mixed 5.85 native vegetation from 3 sites (Beaver Dam, Mesquite and Bunkerville), 18 in tamarisk-dominated vegetation from 3 sites (Big Bend, Desert Springs, Littlefield), and 12 in dead tamarisk-dominated vegetation at 2 sites (Mormon Mesa 1 and 2). We then compared estimated species di-5.90 versity indices in mixed native, tamarisk and dead tamarisk vegetation types using the asymptotic approach for sampling-unit-based incidence data in the program iNEXT (Chao et al. 2016). iNEXT uses an asymptotic approach to infer asymptotic diversity based on statistical estimation 5.95 of the true Hill number at various orders of q (Chao et al. 2014, Chao and Jost 2012). We assessed diversity at 3 orders of q; q = 0 generates an estimate of richness without accounting for relative abundance of species, q = 1 generates an estimate of diversity that incorporates relative fre-5.100 quency of each species and reflects the effective number of common species (analogous to Shannon diversity), and q = 2 generates an estimate that incorporates relative dominance of each species and reflects the effective number of dominant or very abundant species (analogous to 5.105 Simpson's diversity) (Chao et al. 2014). This approach was conservative in that it was based only on whether a species was present at any one point in any one of the 4 visits and 5.106

did not incorporate the number of individuals of a species detected at a point or whether a species was detected in more than 1 count.

We used a similar approach to examine avian responses at smaller spatial scales by testing whether the community of birds detected at points dominated by dead tamarisk in 2013-2014 was different from the community detected at those points before biocontrol in 2009-2010. In this case, detections at individual points characterized as "dead tamarisk" in 2013–2014 were used (n = 12 across 3 sites) and 6.10 compared to the same points sampled in 2009-2010 using the same species diversity estimates at 3 orders of q calculated in iNEXT as above. To control for changes over time independent of tamarisk death, we did the same analysis comparing points characterized in 2013-2014 as "mixed 6.15 native" (13 points over 3 sites). Finally, to determine whether the presence of native vegetation influenced the community of birds detected at points with dead tamarisk, we compared points characterized as "dead tamarisk with some native vegetation" (11 points over 5 sites) in 2013-6.20 2014 using the same approach.

We then assessed whether bird community composition among habitats occupied different community space following biocontrol using an unbiased and unsupervised 6.25 classification approach that is independent of habitat classification. First, we tested the null hypothesis that there are no groupings of bird communities (and therefore the data fit best within one group cluster) using the *factoextra* package (Kassambara and Mundt 2017) for R (R Core Development Team 2018). This analysis assesses the quality 6.30 of group clusters (i.e. how well the data fit within clusters) by calculating the silhouette width for n = 1-6 clusters (i.e. all combinations from a single cluster up to 6 clusters, representing each habitat before and after biocontrol). The silhouette width is a relative measure of confidence for 6.35 group membership within a cluster and values range from -1 to +1 with values closer to 1 represent better clustering (Rousseeuw 1987). Next, we determined the appropriate number of group clusters using package ClValid (Brock et al. 2008) for R which evaluates clustering models and 6.40 the numbers of clustering groups independent of habitat or site classification and subsequently identifies the appropriate clustering algorithm. In our analyses, we evaluated hierarchical, K-means, and partitioning around medoids 6.45 (PAM) clustering models with n = 2-6 clustering groups. *ClValid* assesses group clustering based on three indices: connectivity, Dunn, and silhouette width. The connectivity index assigns group membership of data points based on the spatial proximity to other samples (i.e. points closer in space are assigned to the same group). Connectivity 6.50 ranges from 0 to infinity and smaller values represent well-clustered data (Handl et al. 2005). The Dunn and silhouette indices are measures of the "compactness" and

"spread" of clusters. The Dunn metric is the ratio between 6.55 the smallest distance between data points from different clusters and the largest intracluster distance (Dunn 1974). Dunn indices range from 0 and infinity and higher values represent better clustering. Silhouette values estimate the degree of confidence in membership within a particular 6.60 cluster (Rousseeuw 1987). The silhouette indices are estimated by calculating the mean distance of points within a cluster and the mean distance between clusters and range from -1 to +1 and values close to 1 represent better clustering. Therefore, we chose the number of groups and 6.65 the clustering method in our analyses based on models with optimized connectivity, Dunn, and silhouette values (Brock et al. 2008). In our study, there was no disagreement among indices. Our clustering analysis was optimized at n > 1 clusters, indicating some grouping in bird communities 6.70 (Silhouette value = 0.29, Supplementary Material Table 1) and all clustering indices identified n = 2 clusters and hierarchical clustering as the best algorithm (Supplementary Material Table 1). We then calculated a Euclidean distance matrix among all combinations of community com-6.75 positions in each habitat type before and after biocontrol using the vegan package (Oksanen et al. 2013) for R and AQ9 plotted the results. Finally, we tested for community differences between cluster assignment, biocontrol period, and their interaction using a permutational multivariate 6.80 analysis of variance (PERMANOVA) test with Euclidean distances using the vegan package (Oksanen et al. 2013) for R.

Bird densities. We modeled overall and speciesspecific bird densities (birds ha⁻¹ for each species) using 6.85 the Distance package for program R (R Core Development Team 2018), which accounts for differences in detectability between bird species, habitats, and observers. Speciesspecific densities were calculated for all birds with at least 6.90 25 detections (DeSante 1986). We selected detection functions for each bird species using Akaike's Information Criterion (AIC) and evaluated models using Kolmogorov-Smirnov goodness-of-fit tests (Buckland et al. 2001). For each candidate model, we fit either half-normal or hazard-6.95 rate detection curves and included the covariates site, habitat type, or observer, and additive models including site and observer or habitat type and observer. Because we included covariates, we did not use uniform detection curves (Buckland et al. 2001). Truncation distance for each species was determined by visual inspection of 6.100 detection function plots (Buckland et al. 2001). We then calculated densities (birds ha-1) based on the top model within each site.

Arthropod surveys and microclimate estimates. We assessed differences in the arthropod community by averaging the total number of adult tamarisk beetles, adult tamarisk leafhoppers, adult tamarisk weevils, and all other

6.54

6.106

7.10

S. M. Mahoney et al.

arthropods ("other") collected during each sweep net survey transect and then plotted the mean abundances for group versus habitat type. We then used mixed effects models to test for differences in arthropod abundance between site and habitat types using the *lme4* and *lmerTest* packages in R (R Core Development Team 2018). In our models, arthropod abundance was the response variable, site and habitat type were the fixed effects, and point ID (location of the sample) was included as a random intercept term. We assessed normality and homoscedasticity by visual inspection of QQ and residual plots. We then assessed pairwise differences among habitats using the *lsmeans* package in R. We corrected for multiple hypothesis testing using a Bonferroni correction (corrected $\alpha = 0.015$).

7.15 To assess the relationships between temperature, relative humidity, and tamarisk biocontrol, we determined the maximum temperature and relative humidity for each day from 1 June to 31 July in 2013 and 2014 (Bateman et al. 2013). We then assessed differences among habitats and sites in maximum temperature and relative humidity using 7.20 the *lme4* and *lmerTest* packages in R (R Core Development Team 2018). In our models, maximum daily temperature or relative humidity was the response variable (n = 1 value)day/site for n = 61 days), habitat and site were fixed effects, and datalogger ID number was included as a random inter-7.25 cept term. We assessed normality and homoscedasticity by visual inspection of QQ and residual plots. We then assessed pairwise differences between habitats using the *lsmeans* package in R. We corrected for multiple hypothesis testing using a Bonferroni correction (corrected 7.30 $\alpha = 0.025$).

RESULTS

7.35 Bird Species Diversity, Richness, Community Composition, and Densities

Site-level comparisons of species diversity estimates among habitats characterized as "mixed-native", "tamariskdominated", and "tamarisk-dominated dead" showed that 95% confidence interval (CI) of estimates of effective richness (q = 0) and Shannon diversity (q = 1) overlapped across all sites and times (Figure 2). However, sites characterized as dead in 2013–2014 had lower effective numbers of dominant species (q = 2, Simpson's diversity) both before and after biocontrol (Figure 2).

When the number of sampling points at which each species was detected in each of the 3 vegetation types was compared for communities prior to biocontrol and after biocontrol, only 1 common species (common = detected in >50% of points, n = 10 common species) declined by >30% in the mixed-native sites (Lesser Goldfinch [*Spinus psaltria*]), 3 common species declined by >30% in tamarisk-dominated sites (Brown-headed Cowbird

[Molothrus ater], Song Sparrow [Melospiza melodia], and7.55House Finch [Haemorhous mexicanus]), and 7 common7species declined in dead tamarisk-dominated sites by >30%8(Brown-headed Cowbird, Song Sparrow, Mourning Dove2[Zenaida macroura], Yellow Warbler [Setophaga petechia],7.60Yellowthroat [Geothlypis trichas], and the endangered subspecies Least Bell's Vireo [Vireo bellii pusillus]; Figure 3).7.60

When the community of birds detected at the 12 sampling points characterized as "dead tamarisk" in 2013 werecompared using data collected at the same points both7.65pre- and post-biocontrol, 95% CI overlapped for diversity7.65estimates based on q = 0 (richness) and q = 1 (Shannon7.65diversity) but not for q = 2 (Simpson diversity) indicating7.70that the effective number of dominant species was lower at7.70these points after biocontrol (Figure 4). In contrast, similar7.70comparisons of points characterized as "mixed native" and"dead tamarisk with native component" showed no difference in these parameters between pre- and post-biocontrolcommunities (Figure 4).8

Based on our clustering analyses, all sites grouped to-7.75 gether prior to biocontrol, with the exception of one defoliated tamarisk site (Figure 5). Following biocontrol, mixed habitat sites grouped with the pre-biocontrol cluster and defoliated tamarisk and dead tamarisk sites grouped separately (Figure 5). Bird species composition differed among 7.80 clusters (pseudo- $F_{1,12}$ = 5.68, P < 0.0001), but not between biocontrol periods (pseudo- $F_{1,12} = 1.25$, P = 0.24). However, there was an interaction between biocontrol period and cluster (pseudo- $F_{1,12}$ = 0.13, P = 0.009), indicating community composition in defoliated and dead tamarisk sites 7.85 differed before and after biocontrol, whereas mixed sites remained similar (Figure 5).

Among species that were detected most often (Supplementary Material Table 2) we found several lines of evidence of species declines in tamarisk habitat following biocontrol, based on non-overlapping 95% CIs. Black-chinned Hummingbirds (*Archilochus alexandri*), Least Bell's Vireos, Common Yellowthroats, and Yellow Warblers declined in dead tamarisk sites (Supplementary Material Table 3). House Finches, Lesser Goldfinches, Lucy's Warblers (*Vermivora luciae*), Mourning Doves, Song Sparrows, and Yellow-breasted Chats (*Icteria virens*) declined in defoliated sites following biocontrol (Supplementary Material Table 3).

Some species showed increases in dead tamarisk sites7.100following biocontrol, including Blue-gray Gnatcatchers7.100(Polioptila caerulea), Black-tailed Gnatcatchers8(Polioptila melanura), and Verdins (Auriparus flaviceps,
Supplementary Material Table 3). In defoliated sites,
Black-chinned hummingbirds and Great-tailed Grackles
(Quiscalus mexicanus) increased following biocontrol
(Supplementary Material Table 3).

7.54

7.50

7.40

7.45

7.106

7.90

8.40

8.54

S. M. Mahoney et al.

8.65



FIGURE 2. Site-level diversity estimates and 95% CI at 3 orders of *q* comparing sites characterized as mixed-native in 2009–2010 (open triangles) and 2013–2014 (shaded triangles), tamarisk-dominated in 2009–2010 (open squares) and 2013–2014 (shaded squares), and sites dominated by tamarisk that were largely dead by 2013 in 2009–2010 (open circles) and 2013–2014 (shaded circles). Overall, sites dominated by tamarisk that were largely dead by 2013 had lower effective numbers of dominant species (*q* = 2, Simpson's diversity) both before and after biocontrol. All sites were along the Virgin River in Arizona and Nevada, USA.

Arthropod Surveys and Microclimate Estimates

8.45 Based on mixed effects models, we found no differences in abundances of adult tamarisk beetles ($F_{2,22.6} = 0.92, P = 0.41$) or tamarisk weevils ($F_{2,22.7} = 1.77, P = 0.19$) among habitat types, with leafhoppers ($F_{2,22.4} = 2.93, P = 0.07$) marginally significantly lower in dead tamarisk sites (Supplementary Material Table 4). Non-tamarisk-obligate arthropods, categorized in our study as "other," were more abundant in native mixed sites than in defoliated or dead tamarisk sites (Supplementary Material Table 4, $F_{2,19} = 5.87, P = 0.01$).

We monitored microclimate conditions following biocontrol and from our mixed effects models, daily maximum temperatures increased from 1 June to 31 July $(F_{_{60,840}} = 111.22, P < 0.0001), \text{ but did not vary among sites } (F_{_{2,10}} = 0.51, P = 0.61) \text{ or among habitat type } (F_{_{2,10}} = 1.77, P = 0.22; Figure 6). Relative humidity also increased from 1 June to 31 July (F_{_{60,1740}} = 41.46, P < 0.0001) and varied among habitat types (F_{_{2,25}} = 6.81, P = 0.004; Figure 6), but not among study sites (F_{_{2,25}} = 1.17, P = 0.33; Supplementary Material Supplementary Material Table 5). Humidity in mixed sites was higher than in both defoliated tamarisk and dead tamarisk sites (all post-hoc comparisons <math>P < 0.007;$ Figure 6).

DISCUSSION

Prior to tamarisk biocontrol, bird communities along the Virgin River were similar regardless of the dominant vege-8.70 tation at our study sites. Following biocontrol, however, bird communities in tamarisk habitat that suffered dieback due to biocontrol were different, primarily because of reduced abundance of common species. Our results indicate that when some species, like Yellow Warblers, remain in biocontrol-affected tamarisk stands, they are associated 8.75 with the presence of remnant native cottonwoods and willows. We hypothesize that other species, like Lucy's Warblers and Verdins, may remain because they are able to exploit resources in the surrounding native matrix. Overall, our findings suggest that tamarisk stands during 8.80 the initial recovery from biocontrol dieback could support a moderately diverse assemblage of birds that could act as sources for repopulating these areas if appropriate riparian vegetation establishes after biocontrol. In tamariskdominated areas lacking remnant native vegetation that 8.85 have lost riparian-dependent birds like Yellow Warblers, however, recolonization by those species would need to be through immigration from other stands with a native vegetation component.

8.90 Tamarisk-dominated stands that have experienced extensive dieback as the result of biocontrol can appear to the human eye as unproductive dead zones. Thus, it was surprising that we found no difference between pre- and post-biocontrol in our dead tamarisk sites in the bird com-8.95 munity measures that reflect species richness and Shannon diversity. Although community compositions were significantly different in those dead tamarisk sites pre- and postbiocontrol, primarily due to greater decreases in a broader array of dominant species, the majority of bird species 8.100 were still present, albeit in fewer locations for some species. In terms of species q-diversity estimates (Shannon's and Simpson's diversity calculated from iNEXT), the sites that were characterized as predominantly dead tamarisk in 2013-2014 differed from the other sites in the number of common species both prior to and after biocontrol. Had we 8.105 lacked pre-biocontrol data, the comparison of sites using only post-biocontrol data would have shown a pattern consistent with the hypothesis that these sites differed because 8.106

9.54

S. M. Mahoney et al.



FIGURE 3. More native bird species declined in areas dominated by tamarisk that suffered dieback due to biocontrol. Bars represent the number of census points at which each bird species was detected at sites characterized as mixed-native, tamarisk-dominated, and tamarisk-dominated but largely dead by 2013, prior to biocontrol in 2009–2010 (dark shaded bars) and after biocontrol in 2013–2014 at 8 sites along the Virgin River in Arizona, Nevada, USA. Arrows indicate species in which number of census points at which they were detected declined by at least 30% from 2009–2010 to 2013–2014.

they were largely comprised of dead tamarisk. Instead, those sites already had lower numbers of dominant species prior to biocontrol. Those general community estimates of diversity, however, failed to reflect the compositional 9.40 changes associated with biocontrol and tamarisk death. Based on unbiased clustering algorithms, pre-biocontrol community composition of most sites grouped together, but community composition in mixed and tamariskdominated sites differed following biocontrol, supporting 9.45 the hypothesis that biocontrol altered bird community composition. Some species, like Yellow Warblers, Least Bell's Vireos, Song Sparrows, and Common Yellowthroats, declined markedly in tamarisk-dominated sites that suffered dieback due to defoliation by tamarisk beetles. 9.50

Changes in species abundances were generally consistent with hypotheses proposed by Paxton et al. (2011), in which they predicted 14 species to be sensitive to tamarisk biocontrol due to alterations in prey availability, increased

9.90 nest predation and/or abandonment, or the combined effect of both factors (their Table 1). We found support for this hypothesis in seven species (50%) which showed declines in dead tamarisk sites: densities of Black-chinned Hummingbirds, Least Bell's Vireo, Common Yellowthroats, 9 95 Lucy's Warblers, Mourning Dove, Song Sparrow, and Yellow Warbler decreased. Brown-headed Cowbirds also decreased, possibly in response to reduced host availability. Several species hypothesized by Paxton et al. (2011) to be sensitive to tamarisk biocontrol, including Bewick's 9.100 Wren (Thryomanes bewickii) and Yellow-breasted Chat, showed neutral responses to biocontrol. Our results were generally consistent with a similar study (Darrah and van Riper 2018) that also found a negative relationship between Yellow Warbler densities and tamarisk biocontrol, but con-9.105 trasted in that Song Sparrows decreased in abundance in our study but did not in their study. Interestingly, Verdin abundances increased following biocontrol, possibly 9.106

10 Tamarisk biocontrol and native bird communities

S. M. Mahoney et al.



10.30

10.35

Number of sampling units

FIGURE 4. Survey-point level diversity estimates and 95% Cl at 3 orders of *q* comparing avian communities based on the same points censused pre-biocontrol (oval) and post-biocontrol (triangle) at census points characterized as "mixed native" (*n* = 13), "dead tamarisk with some native component" (*n* = 11) and "dead tamarisk" (*n* = 12). Census points were a subset of those monitored at 8 sites along the Virgin River in Arizona and Nevada, USA, in 2009–2010 pre-biocontrol and in 2013–2014 post-biocontrol. Overall, avian diversity at survey points that included live native vegetation did not differ before or after biocontrol, while diversity was significantly lower at survey points dominated by tamarisk experiencing dieback and death with no live native component.

 because they build enclosed nests (Austin 1970, 1976) that may shield them from microclimate alterations associated with tamarisk biocontrol. Darrah and van Riper (2018)
 found Yellow-breasted Chat densities decreased with tamarisk biocontrol, whereas we found no significant decline. Yellow-breasted Chat densities declined following mechanical removal of tamarisk (Raynor et al. 2017), suggesting that responses to tamarisk removal by species like Yellowbreasted Chat may be more sensitive to habitat structure than whether vegetation is alive or dead.

One hypothesis to explain the changes in the avian community we documented is that tamarisk biocontrol alters the microclimate such that it is too hot or dry for successful nesting (e.g., McLeod 2018, Mueller et al. 2019). Although we did not have pre-biocontrol microclimate estimates, following biocontrol, mean maximum temperatures did not differ among habitats, but temperatures at all sites reached or exceeded temperatures estimated to be lethal for both

10.90 embryos in the egg and adults (approximately >41°C and 47°C, respectively; Lundy 1969, Grant 1982, McKechnie and Wolf 2010). Tamarisk sites had significantly lower humidity, however, and eggs may be particularly sensitive to changes in humidity due to its effect on egg water loss (Deeming 2011). 10.95 Coupled with high temperatures, eggs and adults in tamarisk habitats may lose water more rapidly than in mixed vegetation sites with higher humidity. One caveat of our temperature measurements was that they reflected ambient temperature rather than operative temperature (ambient + solar radiation 10.100 + convection) (Elmore et al. 2017). Given the loss of foliage and increased solar radiation associated with biocontrol, operative temperatures may have been higher in dead tamarisk sites than reflected in our ambient temperature measurements. We know of no studies examining nest microclimate 10.105 in tamarisk-dominated sites that experienced dieback after biocontrol, but studies of Willow Flycatchers at our dead tamarisk sites documented increased rates of nest abandonment 10.106

S. M. Mahoney et al.

Tamarisk biocontrol and native bird communities 11



FIGURE 5. Tamarisk biocontrol significantly altered bird community composition in sites dominated by tamarisk. Dendrogram repre-
sents bird community composition relationships among mixed, defoliated tamarisk (tamarisk*), and tamarisk sites killed by biocontrol
(tamarisk^) before and after tamarisk biocontrol. Tips are labeled with site names. Prior to biocontrol, all but one site (DS) grouped
together in Cluster 1, regardless of habitat type. Following biocontrol, tamarisk-dominated sites grouped in Cluster 2 and mixed vege-
tation sites grouped with Cluster 1, indicating biocontrol altered bird community composition in tamarisk-dominated sites.11.80



FIGURE 6. Tamarisk-dominated sites that experienced biocontrol were hotter (°C, left panel) and drier (%, right panel) than sites with a native vegetation component. Violin plots show distribution of maximum daily temperature and mean maximum daily relative humidity in predominately native, mixed tamarisk-native, predominately defoliated tamarisk (tamarisk*), and dead tamarisk (tamarisk^) habitats after tamarisk biological control. Mean maximum temperatures did not differ among habitats ($F_{2,10} = 1.77$, P = 0.22), but mean relative humidity was higher in mixed and lower in defoliated and dead tamarisk sites ($F_{2,25} = 6.81$, P = 0.004).

Ornithological Applications XX:1–16 © 2022 American Ornithological Society

and the presence of addled eggs with subsequent lower return rates and overall population decline (McLeod 2018).

An alternate hypothesis to explain changes in avian communities after biocontrol is that loss of tamarisk foliage may reduce 12.5 the availability and abundance of insect prey for insectivorous birds (e.g., Pelech and Hannon 1995). Although we did not assess arthropod abundance prior to biocontrol, following biocontrol we found no significant difference in the amount of tamarisk beetles or tamarisk weevils across sites while dead tamarisk 12.10 sites supported marginally lower abundances of tamarisk leafhoppers. Studies along Las Vegas Wash, Nevada, were similar, documenting that tamarisk leafhoppers declined significantly when defoliation reached 60%, while weevils did not (Eckberg and Rice 2016). All 3 tamarisk-obligate insect species have been 12.15 documented in native bird diets. The tamarisk leafhopper has been found to be widely consumed by native birds (Yard et al. 2004, Wiesenborn and Heydon 2007, Durst et al. 2008, Mahoney et al. 2017, van Riper et al. 2018) and the tamarisk weevil was found to be preferred prey for populations of Yellow and Lucy's Warbler along the Virgin River (Mahoney et al. 2017). Non-12.20 tamarisk-obligate arthropods, which would include primarily native species, were significantly more abundant in mixed sites compared to both tamarisk and dead tamarisk sites. Overall, our arthropod comparisons supported the importance of native 12.25 vegetation in maintaining non-tamarisk arthropods, but failed to show a major decline in abundance of tamarisk-obligate insects in the early stages of biocontrol.

Of the 10 insectivorous bird species for which we had enough detections to generate species-specific densities, 3 riparian associates-Least Bell's Vireos, Common 12.30 Yellowthroats, and Yellow Warblers-showed declines following extensive tamarisk defoliation and dieback. Densities of other insectivorous birds may have been maintained by their ability to exploit insect resources 12.35 in the upland habitat surrounding our riparian sites. For example, our tamarisk-defoliated and dead tamarisk sites were largely set within a matrix of mesquite or upland desert, and several species that commonly occur in these upland habitats, Lucy's Warblers, Verdins, and Ash-throated Flycatchers (Myiarchus cinerascens), were 12.40 found in similar densities before and after biocontrol in dead tamarisk sites. Studies of the diet of these species in tamarisk-dominated riparian areas of the Colorado River in Grand Canyon found arthropods in their diet 12.45 more closely reflected the arthropod community of surrounding uplands than that of the tamarisk stands they were captured in (Yard et al. 2004), consistent with the hypothesis that these birds could be maintained in dead tamarisk stands by foraging elsewhere. For other bird spe-12.50 cies, remnant willows or cottonwoods within the tamariskdominated sites may have been critical in allowing them to persist (van Riper et al. 2008). Consistent with this, bird diversity at survey points surrounded by dead tamarisk that also had a willow or cottonwood component were similar12.55before and after biocontrol, whereas points surrounded by
dead tamarisk without native riparian vegetation showed
significantly lower diversity. Likewise, several species were
not recorded in tamarisk-dominated areas following de-
foliation and dieback, including Mourning Doves, Brown-
headed Cowbirds, Song Sparrows, Red-winged Blackbirds,
Common Yellowthroats, Least Bell's Vireos, and Yellow
Warblers, while these species continued to occupy areas
with mixed native vegetation and tamarisk.12.55

Our study documented changes in bird communi-12.65 ties after tamarisk defoliation and dieback that represents the initial stage of recovery following biocontrol using the tamarisk beetle. How long these communitylevel changes will remain and what further changes may 12.70 occur will depend on the longer-term trajectory of plant regeneration after tamarisk dieback. Seedling establishment by cottonwoods and willows is episodic, relying on scouring floods to create suitable germination sites, but if those conditions are met, native riparian vegetation can 12.75 grow rapidly enough to provide habitat for riparian birds relatively quickly. Along the retreating edge of a reservoir in Arizona, for example, native willows and cottonwoods established and grew to a stage that was colonized by Willow Flycatchers in roughly 3 years (Theimer et al. 12.80 2018). Although regeneration of native cottonwoods and willows may occur at these rates in some sites after tamarisk removal, in others it may be slowed or prevented by altered soil characteristics, seed sources, and reduced water availability from human-altered hydrological processes (Stromberg 1998, Stromberg et al. 2009). An analysis 12.85 of over 200 sites across 4 western states at which tamarisk was removed mechanically, chemically or through biocontrol documented that removal enhanced native recovery by <2% over 5 years (González et al. 2017). This suggests 12.90 that at some sites the kinds of changes we documented in avian communities may persist for 5 to 10 years or longer. In other sites, regeneration after defoliation and death may be dominated by other native or non-native plant species. In some of the Virgin River sites we studied, for example, the native shrub arrowweed has increased in abundance 12.95 and overall cover following tamarisk biocontrol (González et al. 2020). Arrowweed is structurally simple compared to willow, cottonwood, and tamarisk, and monotypic stands along the Colorado River attract only a few resident bird species like Mourning Doves, Gambel's Quail, and Abert's 12.100 Towhees (Rosenberg et al. 1991). Thus, areas that become dominated by native species like arrowweed or non-native plant species lacking in structural diversity and insect resources are likely to support bird communities with lower 12.105 richness and abundance (Knutson et al. 2003).

Overall, our results demonstrate that the short-term effects of tamarisk biocontrol on avian communities can be

S. M. Mahoney et al.

S. M. Mahoney et al.

13.55

13.60

Conflict of interest statement: The authors declare no competing interests. Author contributions: All authors conceived of project; S.M.M., M.J.J., and M.K. conducted fieldwork, S.M.M. and T.C.T. analyzed data; S.M.M. and T.C.T. wrote initial manuscript draft; All authors wrote revised paper. Data deposits: Analyses reported in this article can be reproduced using the data provided by Mahoney et al. (2022).

LIIERAIURE CIIED	LITE	ERAT	URE	CITED
------------------	------	------	-----	-------

LITERATURE CITED	13.65
 Anderson, B.W., and R. D. Ohmart (1977). Vegetation structure and bird use in the lower Colorado River Valley. In Importance, Preservation and Management of Riparian Habitat: A Symposium (R. R. Johnson and D. A. Jones, Editors). General Technical Report RM-166, U.S. Department of Agriculture, Forest Service, CO, USA. pp. 23–34. Austin, G. T. (1970). Breeding birds of desert riparian babitat in 	13.70
 Austin, G. T. (1976). Bleeding birds of desert hpanan habitat in southern Nevada. The Condor 72:431–436. Austin, G. T. (1976). Behavioral adaptations of the Verdin to the desert. The Auk 93:245–262. Bateman, H. L., P. L. Nagler, and E. P. Glenn. (2013). Plot-and 	13.75
landscape-level changes in climate and vegetation following defoliation of exotic saltcedar (<i>Tamarix</i> sp.) from the biocon- trol agent <i>Diorhabda carinulata</i> along a stream in the Mojave Desert (USA). Journal of Arid Environments 89:16–20.	
Bean, D. W., I. Dudley, and K. Hultine (2013). Bring on the bee- tles! The history and impact of tamarisk biological control. In Tamarix: A Case Study of Ecological Change in the American West (A. Sher, and M. F. Quigley, Editors). Oxford University Press, Oxford, UK. pp. 377–403.	13.80
Brand, L. A., G. C. White, and B. R. Noon (2008). Factors influencing species richness and community composition of breeding birds in a desert riparian corridor. The Condor 110:199–210.	13.85
Brock, G., V. Pihur, S. Datta, and S. Datta (2008). clValid: An R package for cluster validation. Journal of Statistical Software 25:1–22.	
 Buckland, S. T., D. R. Anderson, K. P. Burnham, J. L. Laake, D. L. Borchers, and L. Thomas (2001). Introduction to Distance Sampling. Oxford University Press, Oxford, UK. Cable T. T. W. H. Fick, and F. J. Baynor (2015). The potential im- 	13.90
pacts of salt cedar eradication (Tamarix sp.) on the birds of the Cimarron National Grassland. Transactions of the Kansas Academy of Science 118:41–47.	13.95
 Cartron, J-L. E., S. H. Stoleson, and R. R. Jonnson (1999). Riparian dependence, biogeographic status, and likelihood of endangerment of landbirds of the Southwest. In Rio Grande Ecosystems: Linking Land, Water, and People (D. M. Finch, J. C. Whitney, J. F. Kelly, and S. R. Loftin, Editors). Proceedings RMRS-P-7, U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station. Ogden, UT, USA, pp. 211–215. 	13.100
Chao, A., and L. Jost (2012). Coverage-based rarefaction and ex- trapolation: standardizing samples by completeness rather than size. Ecology 93:2533–2547.	
Chao, A., N. J. Gotelli, T. C. Hsieh, E. L. Sander, K. H. Ma, R. K. Colwell, and A. M. Ellison (2014). Rarefaction and extrapolation with Hill numbers: A framework for sampling and estimation in spe-	13.105
cies diversity studies. Ecological Monographs 84:45–67.	13.106

	substantial in sites already dominated by tamarisk, in that
	dieback will have greater relative impact to overall habitat
	suitability than in ecosystems where significant native ri-
	parian vegetation remains. How long these effects remain
13.5	will vary depending upon the rate of native plant recovery,
	and therefore we recommend monitoring sites to deter-
	mine the trajectory of native recovery over time. If native
	tree recovery in these sites is rapid, avian communities
	may recover rapidly as well. However, in areas where na-
13.10	tive tree recovery is slow or forestalled by invasion by other
	non-native plant species, land managers will need to con-
	sider the feasibility of active restoration, which can be cost-
	prohibitive (Taylor and McDaniel 1998), particularly where
	the altered hydrologic regimes of southwest rivers may
13.15	limit the ability of native cottonwoods and willows to ger-
	minate (Stromberg 1998, Stromberg et al. 2009). Therefore,
	restoration efforts should assess the likelihood of success
	based on an evaluation of several factors including water
	availability necessary for native vegetation establishment
13.20	(Stromberg 1998), the abundance and distribution of ex-
	tant native vegetation, the potential impacts of flooding
	and fire, and financial resources available (Shafroth et al.
	2013, Orr et al. 2014). More broadly, our study highlights
	the ability of many bird species to remain in areas greatly
13.25	modified by invasion and subsequent biocontrol, and the
	importance of even relatively small remnants of native
	vegetation in allowing species sensitive to those changes
	to persist.

13.30 SUPPLEMENTARY MATERIAL

Supplementary material is available at Ornithological Applications online.

13.35

13.54

ACKNOWLEDGMENTS

	We offer our sincerest gratitude to K. Becraft, P. Costa,
	U. Kall, J. Kreitzer, and J. Agee, D. N. Kakestraw, and Z. D.
13.40	Watson for their field and lab assistance. We thank R. L.
	Hammond and B. J. Butterfield for their guidance with
	statistical analyses, and C. E. Aslan, B. Pasch, and S. M.
	Shuster for reviewing early versions of this manuscript.
	Preliminary data were presented at several RiversEdge
12 45	West Conferences and discussions during the meetings
15.45	were critical for the final version of this paper. Comments
	from C. A. Lindell, E. Blomberg, and two anonymous re-
	viewers substantially improved this manuscript.
	Funding statement: We would like to acknowledge the
13.50	Landscape Conservation Cooperative (LCC), Clark County
	Multiple Species Habitat Conservation Plan (MHSCP), and
	the United States Department of Interior (USDI) for their fi-
	nancial support for this project.

Ethics statement: All work was approved by institutional IACUC, BLM, and USFWS.

Ornithological Applications XX:1-16 © 2022 American Ornithological Society

14.25

14.35

14.45

14.54

Tamarisk biocontrol and native bird communities 14

S. M. Mahoney et al.

- Chao, A., K. H. Ma, and T. C. Hsieh (2016). iNEXT (iNterpolation and EXTrapolation) Online: Software for Interpolation and Extrapolation of Species Diversity. Program and User's Guide. http://chao.stat.nthu.edu.tw/wordpress/software_download/
- Darrah, A. J., and C. van Riper III (2018). Riparian bird density decline in response to biocontrol of Tamarix from riparian ecosystems along the Dolores River in SW Colorado, USA. Biological Invasions 20:709-720.

Deeming, D. C. (2011) Importance of nest type on the regulation of humidity in bird nests. Avian Biology Research 4:23–31.

- DeLoach, C. J., R. I. Carruthers, J. E. Lovich, T. L. Dudley, and 14.10S. D. Smith (2000). Ecological interactions in the biological control of saltcedar (Tamarix spp.) in the United States: Toward a new understanding. In Proceedings of the X International Symposium on Biological Control of Weeds (N. R. Spencer, Editor). Montana State University, Bozeman, Montana, USA. pp. 819-8731. 14.15
 - DeSante, D. F. (1986). A field test of the variable circular-plot censusing method in a Sierran subalpine forest habitat. The Condor 88:129–142.
- Dudley, T. L., and D. W. Bean (2012). Tamarisk biocontrol, endangered species risk and resolution of conflict through riparian 14.20 restoration. BioControl 57:331-347.
 - Dunn, J. C. (1974). Well-separated clusters and optimal fuzzy partitions. Journal of Cybernetics 4:95–104.
 - Durst, S. L., T. C. Theimer, E. H. Paxton, and M. K. Sogge (2008). Temporal variation in the arthropod community of desert riparian habitats with varying amounts of saltcedar (Tamarix ramosissima). Journal of Arid Environments 72:1644–1653.
 - Eckberg, J. R., and N. A. Rice (2016). Northern tamarisk beetle (Diorhabda carinulata) effects on established tamarisk-feeding invertebrate populations along the Las Vegas Wash, Clark County, Nevada. The Southwestern Naturalist 61:101–107.
- Ellis, L. M. (1995). Bird use of saltcedar and cottonwood vegetation 14.30 in the Middle Rio Grande Valley of New Mexico, USA. Journal of Arid Environments 30:339-349.
 - Elmore, R. D., J. M. Carroll, E. P. Tanner, T. J. Hovick, B. A. Grisham, S. D. Fuhlendorf, and S. K. Windels (2017). Implications of the thermal environment for terrestrial wildlife management. Wildlife Society Bulletin 41:183-193.
 - Fleishman, E., N. McDonal, R. M. Nally, D. D. Murphy, J. Walters, and T. Floyd (2003). Effects of floristics, physiognomy and non-native vegetation on riparian bird communities in a Mojave Desert watershed. Journal of Animal Ecology 72:484-490.
- 14.40Friedman, J. M., G. T. Auble, P. B. Shafroth, M. L. Scott, M. F. Merigliano, M. D. Freehling, and E. R. Griffin (2005). Dominance of non-native riparian trees in western USA. Biological Invasions 7:747-751.
 - Gaskin, J. F., and B. A. Schaal (2002). Hybrid Tamarix widespread in U.S. invasion and undetected in native Asian range. Proceedings of the National Academy of Sciences of the United States of America 99:11256-11259.
- González, E., P. B. Shafroth, S. R. Lee, S. M. Ostoja, and M. L. Brooks (2020). Combined effects of biological control of an invasive shrub and fluvial processes on riparian vegetation dynamics. 14.50Biological Invasions 22:2339–2356.
 - González, E., A. A. Sher, R. M. Anderson, R. F. Bay, D. W. Bean, G. J. Bissonnete, B. Bourgeois, D. J. Cooper, K. Dohrenwend, K. D. Eichhorst, and H. El Waer (2017). Vegetation response to invasive Tamarix control in southwestern US

rivers: A collaborative study including 416 sites. Ecological 14.55 Applications 27:1789-1804.

- Grant, G. S. (1982). Avian incubation: Egg temperature, nest humidity, and behavioral thermoregulation in a hot environment. Ornithological Monographs, no. 30. American Ornithologists' Union, Washington, D.C., USA.
- 14.60 Handl, J., J. Knowles, and D. B. Kell (2005). Computational cluster validation in post-genomic data analysis. Bioinformatics (Oxford, England) 21:3201-3212.
- Hobbs, R. J., E. Higgs, and J. A. Harris (2009). Novel ecosystems: implications for conservation and restoration. Trends in Ecology & Evolution 24:599-605.

Hobbs, R. J., L. E. Valentine, R. J. Standish, and S. T. Jackson (2018). Movers and Stayers: Novel assemblages in changing environments. Trends in Ecology & Evolution 33:116-128.

- Holmes, J. A., J. R. Spence, and M. K. Sogge (2005). Birds of the Colorado River in Grand Canyon: A synthesis of status, trends, and dam operation effects. In The State of the Colorado 14.70 River Ecosystem in Grand Canyon (S. P. Glass, J. E. Lovich, and T. E. Melis, Editors). U.S. Geological Survey Circular 128, Federal Center, Denver, CO, USA. pp. 123-138.
- Howlett, J. S., and B. J. Stutchbury (1996). Nest concealment and predation in Hooded Warblers: Experimental removal of nest cover. The Auk 113:1-9.
- Hultine, K. R., T. L. Dudley, D. F. Koepke, D. W. Bean, E. P. Glenn, and A. M. Lambert (2015). Patterns of herbivory-induced mortality of a dominant non-native tree/shrub (Tamarix spp.) in a southwestern US watershed. Biological Invasions 17:1729-1742.
- Hunter, W.C., R. D. Ohmart, and B. W. Anderson (1988). Use of exotic saltcedar (Tamarix chinensis) by birds in arid riparian systems. The Condor 90:113-123.
- Johnson, R.R., L. T. Haight, and J. M. Simpson (1977). Endangered species vs. endangered habitats: a concept. In Importance, Preservation and Management of Riparian Habitat: A Symposium (R. R. Johnson and D. A Jones, Editors). General Technical Report RM 43, U.S. Department of Agriculture, Forest Service, Fort Collins, CO, USA. pp. 68-74.
- Kassambara, A., and F. Mundt (2017). Factoextra: Extract and visualize the results of multivariate data analyses. R Package Version 1:337–354. https://cran.r-project.org/web/packages/ factoextra/readme/README.html
- Knutson, A. E., M. A. Muegge, T. O. Robbins, and C. J. DeLoach (2003). Insects associated with saltcedar, baccharis and willow in west Texas and their value as food for insectivorous birds, preliminary results. In Saltcedar and Water Resources in the West. Texas Agricultural Experiment Station, San Angelo, Texas, USA. pp. 38-47.
- Lewis, P. A., C. J. deLoach, A. E. Knutson, J. L. Tracy, and T. O Robbin (2003). Biology of Diorhabda elongata deserticola (Coleoptera: Chrysomelidae), an Asian leaf beetle for biological control of saltcedars (Tamarix spp.) in the United States. Biological Control 27:101-116.
- Longland, W., and T. Dudley (2008). Effects of a biological control agent on the use of saltcedar habitat by passerine birds. Great Basin Birds 10:21–26.
- Lundy, H (1969). A review of the effects of temperature, humidity, turning, and gaseous environment in the incubator on the 14.105 hatchability of the hen's egg. In The Fertility and Hatchability of the Hen's Egg (T. C. Carter and B. M. Freeman, Editors). Oliver and Boyd, Edinburgh, UK. pp. 143–176. 14.106

AQ1014.75

14.65

14.80

14.85

14.95

14.100

S. M. Mahoney et al.

- Mahoney, S. M., M. J. Johnson, J. A. Holmes, T. L. Dudley, M. Kuehn, and T. C. Theimer (2022). Data from: Tamarisk biocontrol alters bird community composition in the absence of cottonwood and willow vegetation. Ornithological Applications 124:duac000. doi:10.5061/dryad.vt4b8gttw.
- 15.5^{AQ11} Mahoney, S. M., T. C. Theimer, M. J. Johnson, and J. T. Foster (2017). Similar dietary but different numeric responses to nonnative tamarisk (Tamarix spp.) by two native warblers. Biological Invasions 19:1935–1950.
 - Mahoney, S. M., R. R. Winton, L. M. McCabe, and M. J. Johnson (2020). New locality for the biological control agent (Diorhabda spp.) in Northern Arizona. Southwestern Entomologist 45:293-296.
 - McLeod, M. A. (2018). Unintended consequences: Tamarisk control and increasing threats to the southwestern willow flycatcher. In Riparian Research and Management: Past, Present,
 - Future: Volume 1 (R. R. Johnson, S. W. Carothers, D. M. Finch, K. J. Kingsley, and J. T. Stanley, Editors). General Technical Report RMRS-GTR-377, U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station, Fort Collins, CO, USA. pp. 62-84.
 - Mosher, K. R., and H. L. Bateman (2016). The effects of riparian restoration following saltcedar (Tamarix spp.) biocontrol on habitat and herpetofauna along a desert stream. Restoration Ecology 24:71-80.
 - Mueller, A. J., K. D. Miller, and E. K. Bowers (2019). Nest microclimate during incubation affects posthatching development and parental care in wild birds. Scientific Reports 9: 5161.
 - Nagler, P.L., T. Brown, K. R. Hultine, C. van Riper III, D. W. Bean, P. E. Dennison, R. S. Murray, and E. P. Glenn (2012). Regional scale impacts of Tamarix leaf beetles (Diorhabda carinulata) on the water availability of western U.S. Rivers as determined by multi-scale remote sensing methods. Remote Sensing of Environment 118:227-240.
 - Nagler, P. L., U. Nguyen, H. L. Bateman, C. J. Jarchow, E. P. Glenn, W. J. Waugh, and C. van Riper III (2018). Northern tamarisk beetle (Diorhabda carinulata) and tamarisk (Tamarix spp.) interactions in the Colorado River Basin. Restoration Ecology 26:348-359
 - Orr, B., M. Johnson, G. Leverich, T. Dudley, J. Hatten, Z. Diggory, K. Hultine, D. Orr, and S. Stone (2017). Multi-scale riparian restoration planning and implementation on the Virgin and Gila Rivers. In Case Studies of Riparian and Watershed Restoration Areas in the SW U.S.—Principles, Challenges, and Successes
 - (B. E. Ralston, D. A. Sarr, Editors). USGS Open File Report 2017-1091, U.S. Department of the Interior, U.S. Geological Survey, Reston, VA, USA.
 - Paxton, E. H., T. C. Theimer, and M. K. Sogge (2011). Tamarisk biocontrol using tamarisk beetles: Potential consequences for riparian birds in the southwestern United States. The Condor 113:255-265.
 - Pelech, S., and S. J. Hannon (1995). Impact of tent caterpillar defoliation on the reproductive success of Black-capped Chickadees. The Condor 97:1071-1074.
 - Pearson, D. E., and R. M. Callaway (2003). Indirect effects of host-specific biological control agents. Trends in Ecology & Evolution 18:456-461.
 - R Core Development Team (2018). The R Project for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. https://www.R-project.org/

- Raynor, E. J., T. T. Cable, and B. K. Sandercock (2017). Effects of 15.55 *Tamarix* removal on the community dynamics of riparian birds in a semiarid grassland. Restoration Ecology 25:778-787.
- Reynolds, R. T., J. M. Scott, and R. A. Nussbaum (1980). A variable circular-plot method for estimating bird numbers. The Condor 82:309-313.
- Rodriguez, L. F. (2006). Can invasive species facilitate native species? Evidence of how, when, and why these impacts occur. Biological Invasions 8:927–939.
- Rosenberg, K. V., R. D. Ohmart, W. C. Hunter, and B. W. Anderson (1991). Birds of the Lower Colorado River Valley. University of Arizona Press, Tucson, AZ, USA.
- Rousseeuw, P. J. (1987). Silhouettes: A graphical aid to the interpretation and validation of cluster analysis. Journal of Computational and Applied Mathematics 20:53-65.
- Shafroth, P. B., J. R. Cleverly, T. L. Dudley, J. P. Taylor, C. van Riper, 3rd, E. P. Weeks, and J. N. Stuart (2005). Control of Tamarix in the Western United States: implications for water salvage, wild-15.70 life use, and riparian restoration. Environmental Management 35:231-246.
- Shafroth, P. B., D. M. Merritt, M. K. Briggs, V. B. Beauchamp, K. D. Lair, M. L. Scott, and A. A. Sher (2013). Riparian restoration in the context of Tamarix control. In Tamarix: A Case Study of Ecological Change in the American West (A. Sher, and M. F. Quigley, Editors). Oxford University Press, Oxford, UK. pp. 404-425.
- Skagen, S. K., C. P. Melcher, W. H. Howe, and F. L. Knopf (1998). Comparative use of riparian corridors and oases by migrating birds in southeast Arizona. Conservation Biology 12:896-909.
- Smith, A. N., M. J. Anderson, and M. D. Pawley (2017). Could ecologists be more random? Straightforward alternatives to haphazard spatial sampling. Ecography 40:1251-1255.
- Sogge, M. K., S. J. Sferra, and E. H. Paxton (2008). Tamarix as habitat for birds: Implications for riparian restoration in the southwestern United States. Restoration Ecology 16:146–154.
- Sogge, M. K., E. H. Paxton, and C. van Riper III (2013). Tamarisk in riparian woodlands: A bird's eye view. In Tamarix: A Case Study of Ecological Change in the American West (A. Sher and M. F. Quigley, Editors). Oxford University Press, Oxford, UK. pp. 189-206.
- Stevens, L. E., B. T. Brown, J. M. Simpson, and R. R. Johnson (1977). The importance of riparian habitat to migrating birds. In Importance, Preservation and Management of Riparian Habitat: A Symposium. General Technical Report RM-43, U.S. Department of Agriculture, Forest Service, Tucson, AZ, USA. pp. 156-164.
- Stromberg, J. (1998). Dynamics of Fremont cottonwood (Populus fremontii) and saltcedar (Tamarix chinensis) populations along the San Pedro River, Arizona. Journal of Arid Environments 40:133-155.
- Stromberg, J. C., M. K. Chew, P. L. Nagler, and E. P. Glenn (2009). 15.100 Changing perceptions of change: The role of scientists in Tamarix and river management. Restoration Ecology 17:177-186.
- Strong, T. R., and C. E. Bock (1990). Bird species distribution patterns in riparian habitats in southeastern Arizona. The Condor 92:866-885.
- Taylor, J. P., and K. C. McDaniel (1998). Restoration of saltcedar (Tamarix spp.)-infested floodplains on the Bosque del Apache National Wildlife Refuge. Weed Technology 12:345–352.

15.106

15.105

15.60

15.65

15.75

15.80

15.85

15.90

15.95

- 15.10

15.15

15.20

15.25

15.30

15.35

15.40

15.45

15.50

	16 Tamarisk biocontrol and native bird communities	S. M. Mahoney et al.	
	Theimer, T. C., M. K. Sogge, S. N. Cardinal, S. L. Durst, and E. H. Paxton (2018). Extreme drought alters frequency and re- productive success of floaters in Willow Flycatchers. The Auk: Ornithological Advances 135:647–656.	diets along the Dolores River in Southwestern Colorado USA. Biological Invasions 20:3145–3159. Vilà, M., J. L. Espinar, M. Hejda, P. E. Hulme, V. Jarošík, J. L. Maron, J. Pergl, U. Schaffner, Y. Sun, and P. Pyšek (2011). Ecological im- pacts of invasive alien plants: a mota analysis of their effects.	16.55
16.5	biological control of saltcedar (Tamarix spp.) in thirteen states. Environmental assessment. Animal and Plant Health Inspection Service, USDA, Western Region 2. https://www. aphis.usda.gov/plant_health/ea/downloads/salteafonsi.	 pacts of invasive alien plants: a meta-analysis of their effects on species, communities and ecosystems. Ecology Letters 14:702–708. Wiesenborn, W. D., and S. L. Heydon (2007). Diets of breeding Southwestern Willow Flycatchers in different habitats. The Wilson Journal of Ornithology 119:547–558. 	16.60
16.10	 van Riper, C. III, K. L. Paxton, C. O'Brien, P. B. Shafroth, and L. J. McGrath (2008). Rethinking avian response to <i>Tamarix</i> on the lower Colorado River: A threshold hypothesis. Restoration Ecology 16:155–167. van Riper, C. III, S. L. Puckett, and A. J. Darrah (2018). Influences of the invasive tamarisk beetle (<i>Diorhabda carinulata</i>) on avian 	 Yamaura, Y., F. G. Blanchet, and M. Higa (2019). Analyzing community structure subject to incomplete sampling: hierarchical community model vs. canonical ordinations. Ecology 100:e02759. Yard, H. K., C. van Riper III, B. T. Brown, and M. J. Kearsley (2004). Diets of insectivorous birds along the Colorado River in Grand Canyon, Arizona. The Condor 106:106–115. 	16.65
16.15			16.70
16.20			16.75
16.25			16.80
16.30			16.85
16.35			16.90
16.40			16.95
16.45			16.100
16.50			16.105
16.54			16.106